

Hydrogen Sulfide Is a Signaling Molecule and a Cytoprotectant

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Abstract

Significance: Accumulating evidence shows that hydrogen sulfide may function as a signaling molecule in processes such as neuromodulation in the brain and smooth muscle relaxation in the vascular system. It also has a cytoprotective effect, since it can protect neurons and cardiac muscle from oxidative stress and ischemia-reperfusion injury, respectively. Hydrogen sulfide can also modulate inflammation, insulin release, and angiogenesis. **Recent Advances:** The regulation of the activity of 3-mercaptopyruvate sulfur transferase (3MST) along with cysteine aminotransferase (CAT), one of the H₂S producing pathways, has been demonstrated. The production of H₂S by the pathway, which is regulated by Ca²⁺ and facilitated by thioredoxin and dihydrolipoic acid, is also involved in H₂S signaling as well as cytoprotection. Sulfur hydration of proteins by H₂S has been proposed to modulate protein functions. H₂S-sensitive fluorescent probes, which enable us to measure the localization of H₂S in real time, have been developed. **Critical Issues:** The basal concentrations of H₂S have recently been measured and found to be much lower than those initially reported. However, the concentration of H₂S reached in stimulated cells, as well as the regulation of H₂S producing enzymes is not well understood. It has been proposed that some of the effects of H₂S on the regulation of enzymes and receptors might be explained through the properties of sulfane sulfur (S⁰), another form of active sulfur. **Future Directions:** The determination of H₂S concentrations in activated cells using new methods including H₂S-sensitive fluorescent probes, as well as the investigation of the effects of H₂S using specific inhibitors, may provide better understanding of the physiological function of this molecule. Clarifying mechanisms of H₂S activity may also facilitate the development of new therapeutic compounds. *Antioxid. Redox Signal.* 17, 45–57.

Introduction

THE ENDOGENOUS CONCENTRATIONS OF SULFIDE in the brains of humans, bovine, and rats are in the range of 50–160 μ M, strongly suggesting that endogenous H₂S may have significant physiological functions (20, 56, 72). However, the concentrations of sulfide measured in these studies, which contained H₂S released from acid-labile sulfur in addition to free H₂S, were overestimated (23). Recently, the basal concentrations of H₂S in tissues have been measured and found to be much lower than those previously reported (18, 24, 36). During the preparation of this manuscript, H₂S-sensitive fluorescence probes have been reported from several laboratories (38, 39, 50, 52, 55). These probes may enable us to detect the sites at which H₂S is released in real time and assess the local concentrations of H₂S.

We demonstrated that cystathionine β -synthase (CBS), which produces H₂S from cysteine or cysteine with homocysteine, is localized to astrocytes, a type of glia, and 3-mercaptopyruvate sulfur transferase (3MST), which produces

H₂S from 3-mercaptopyruvate, is localized to neurons in the brain (16, 57). The activity of CBS is enhanced by S-adenosyl methionine, and that of 3MST pathway is regulated by Ca²⁺ (1, 43). Cystathionine γ -lyase (CSE) is expressed little in the brain (1, 26).

H₂S facilitates the induction of hippocampal long-term potentiation by enhancing the activity of N-methyl D,L-aspartate (NMDA) receptors (1). In the hypothalamus, H₂S suppresses the KCl-stimulated release of corticotropin-releasing hormone (CRH), while it has no effect on the basal release of CRH. S-Adenosyl methionine inhibits the release of glucocorticoid induced by stress without any effect on hypothalamo-pituitary-adrenal function under resting conditions *in vivo* (13). H₂S induces Ca²⁺ influx in astrocytes that propagates to the surrounding astrocytes as Ca²⁺ waves (16, 47). There is a reciprocal interaction between neurons and astrocytes: neuronal excitation activates surrounding astrocytes, which in turn modulate synaptic activity. H₂S regulates synaptic activity by enhancing the activity of both neurons and astrocytes.

In addition to a possible function as a signaling molecule, we demonstrated a role of H_2S as a cytoprotectant in the brain (30–32). H_2S protects neurons from oxidative stress by restoring glutathione levels decreased by oxidative stress and suppressing oxidative stress in mitochondria. Philip Moore and colleagues also demonstrated that H_2S protects a neuroblastoma cell line from oxidative stress induced by peroxynitrite and hypochlorous acid (76, 77). These oxidants cause the oxidative modification of proteins and lipids seen *in vivo* in patients with neurodegenerative diseases such as Alzheimer's disease and Parkinson's disease. H_2S also protects retinal neurons from light-induced degeneration by suppressing the excessive Ca^{2+} influx (43). The cytoprotective effect was also found by David Lefer and colleagues in the cardiovascular system. H_2S protects cardiac muscle from ischemia-reperfusion injury by preserving mitochondrial function (15). The application of an H_2S donor or the regulation of endogenous H_2S concentrations by manipulating the CSE gene protected cardiomyocytes from ischemia-reperfusion injury. Based on this study, an H_2S donor is being given to the bypass surgery patients in a Phase II clinical trial (51).

We showed that CSE is expressed in the thoracic aorta, portal veins, and ileum, and that H_2S relaxes these tissues (22). The smooth muscle relaxant effect of H_2S in the thoracic aorta and ileum was further studied by Rui Wang and Philip Moore, and H_2S was found to activate adenosine triphosphate (ATP)-dependent K^+ (K_{ATP}) channels (65, 82). The role of H_2S and CSE has also been studied *in vivo* using CSE-knockout mice (25, 81). The regulation of inflammation, insulin release, and angiogenesis by H_2S have also been intensively studied, and this has led to the development of therapeutic agents. This review focuses on the regulation of endogenous H_2S and the regulation of its enzymatic production, as well as the methods recently developed to measure it in addition to the function as a signaling molecule and a cytoprotectant.

Permeability of H_2S Through the Plasma Membrane

The mechanism of H_2S transport through the plasma membrane has recently been demonstrated. Because of the structural similarity of H_2S with H_2O , it was hypothesized that H_2S may pass through aquaporins or water channels (40). However, although reconstituted aquaporin obtained from a sulfide-reducing bacteria, AfaAQP, increased the permeability of water in planar lipid bilayers, the permeability of H_2S remained unchanged. The membrane resistance to H_2S permeation is negligible. Even bilayers containing cholesterol and sphingomyelin, which decrease the diffusiveness of the membrane, had the same H_2S permeability. These observations indicate that the membrane is not the limiting factor for H_2S transport (40) (Fig. 1). The aquaporin homolog AqpM, which is derived from bacteria that rely on H_2S as the terminal electron acceptor for energy production remains a candidate H_2S transporter (34).

H_2S is soluble in water and dissociates into H^+ and HS^- . In physiologic saline at 37°C and pH 7.4, approximately one-fifth of H_2S exists as the undissociated form (H_2S), and the remaining four-fifths exists as HS^- plus a trace of S^{2-} at equilibrium with H_2S (14). However, theoretical calculations indicate that transport of HS^- by Cl^- channels or other anion channels is unlikely to play a physiological role (40). The form of the molecule passing through the membrane is not neces-

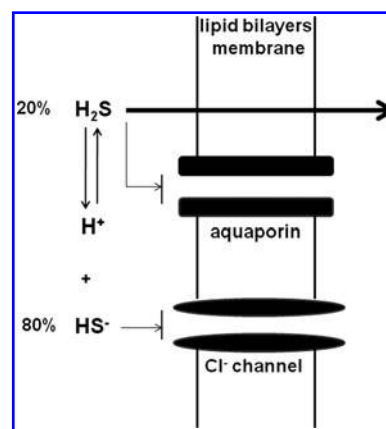


FIG. 1. H_2S passes through the plasma membrane by diffusion. Aquaporin and anion channels are not required for H_2S and HS^- respectively to pass through the plasma membrane.

sarily the functional form. It may be difficult to determine which form, H_2S or HS^- , has a physiological function. In addition, since HS^- can be oxidized to persulfide, it is possible that some observed effects with NaHS or Na_2S may be even induced by a trace amount of persulfide. As to the possible involvement of other sulfur-containing substances as signaling molecules, see the review by Toohey (68).

H_2S Concentrations

In early studies, the concentrations of sulfide in the brain were measured under conditions with high concentrations of acids. Therefore, H_2S released from acid-labile sulfur was measured along with free H_2S , with a combined concentration of between 50–160 μM in the brain (20, 56, 72). Acid-labile sulfur is derived from sulfur atoms in the iron–sulfur center of mitochondria respiratory chain enzymes. A pH less than 5.4 releases H_2S from this source of acid-labile sulfur (24). Since mitochondrial pH is between 7 and 8, H_2S may not be released from acid-labile sulfur under physiologic conditions, and therefore, the reported intracellular concentrations for H_2S are likely overestimated.

The concentrations of free H_2S are much less than those of acid-labile sulfur. Brain homogenates were vigorously mixed in the syringe, and released H_2S was measured by gas chromatography. By this method, the concentration of free H_2S is 14 nM (0.12 $\mu\text{mol/kg}$ protein), while that released from acid-labile sulfur is 916 $\mu\text{mol/kg}$ protein (18, 36). In another method, free H_2S is below detectable levels, while that of H_2S released from acid-labile sulfur is 161 nmol/g protein (24). Although 1 μM Na_2S efficiently activates vacuolar-type H^+ ATPase to suppress Ca^{2+} influx in the retina, the effects of the basal levels of H_2S (10–20 nM) have not been reported. In order to exert its physiological function, μM concentrations of H_2S have to be released from the activated enzymes or the intracellular store, but it is enough in a short time of period in a restricted area. After the induction of responses, H_2S may be re-stored or degraded to be a steady state to cease the reaction. The basal concentrations of H_2S are maintained in low levels in cells to properly respond to H_2S . For example, the application of high concentrations of H_2S (200 μM NaHS) to astrocytes suppresses Ca^{2+} influx induced by the second

application, suggesting that high concentrations of H₂S cause desensitization to H₂S (47).

More recent measurements of the basal concentrations of H₂S in blood still exhibit significant variation. For example, free H₂S in blood measured by gas chromatography, was 0.07 μ mol/kg protein (approximately 8 nM) (36). In this method, the efficiency of recovering H₂S from a control sodium sulfide solution was estimated as a blank value in the absence of tissue homogenates. However, tissue homogenates efficiently absorb H₂S (24), and it is therefore possible that the blank value may be overestimated compared to that without tissues, resulting in the relatively lower estimate of free H₂S values in tissue homogenates. Another method mixed blood with monobromobimane, the thiol-specific derivatization agent, and recovered thiol-bound monobromobimane. The concentrations of H₂S-bound monobromobimane were specifically measured by mass spectrometry (79). Free H₂S in blood with this method was in the range of 0.4–0.9 μ M. Depending on the method used, the concentration of free H₂S vary up to 100-fold, but a consistent finding is that the basal free H₂S in blood is maintained at a low concentration.

A low basal concentration of H₂S does not necessarily indicate that these are the biologically functional concentrations of this molecule. Because the physiological stimuli of H₂S release have not been identified, it is difficult to estimate the local concentrations of H₂S reached when cells are activated. Additionally, the regulation of the activity of H₂S-producing enzymes is also not well understood. The production of H₂S by CBS is enhanced by S-adenosyl methionine two-fold compared to control samples (1). Activity of the 3MST/CAT pathway is regulated by Ca²⁺. The activity is highest in the absence of Ca²⁺ and suppressed by Ca²⁺ in a concentration-dependent manner. The production of H₂S by this pathway is dramatically changed in the range of intracellular Ca²⁺ (43). Although the regulation of CSE by Ca²⁺/calmodulin was reported, the concentrations of Ca²⁺ applied in the study were supraphysiological (1–2 mM), as the intracellular concentrations of Ca²⁺ (100 nM–3 μ M) (81).

Another source of H₂S is bound sulfane sulfur, which may function as an intracellular store of H₂S. Cells expressing 3MST and CAT contain twice as much bound sulfane sulfur as control cells. The concentrations of bound sulfane sulfur are low in cells expressing a mutant 3MST that lacks H₂S-producing activity (57). Lysates of neurons and astrocytes release H₂S in the presence of endogenous levels of glutathione and cysteine at pH 8.4, and brain homogenates release it in the presence of dihydrolipoic acid (DHLA) at pH 8.0 (24, 42). H₂S can be released from astrocytes by high K⁺, which is induced by neuronal excitation (24). The concentration of bound sulfane sulfur in the brain is 1500 nmol/g protein, which is sufficient to release H₂S to stimulate target molecules (24). Considering the fact that the K_m values of H₂S-producing enzymes for substrates are greater than the endogenous concentrations, the endogenous production rate of H₂S must be very low. Therefore, it is possible that H₂S may be stored after it is produced by enzymes, and released from its intracellular stores when cells are stimulated. The percentage of bound sulfane sulfur that releases H₂S in response to cell activation has not been determined (Fig. 2). It is critical that the local concentrations of H₂S reached upon cell stimulation are now re-quantified using methods including H₂S sensitive probes (38, 39, 50, 52, 55).

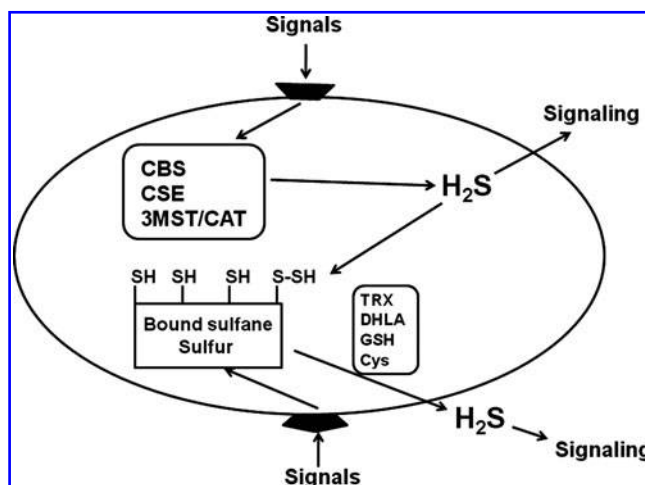


FIG. 2. H₂S is released from the producing enzymes and bound sulfane sulfur. H₂S produced by enzymes may function as a signaling molecule and may also be stored in bound sulfane sulfur as intracellular stores. When cells are stimulated, H₂S can be released from the stores. Although lysates of neurons and astrocytes release H₂S in the presence of endogenous reducing substances, H₂S release from intact cells with physiological stimulations has not been fully elucidated. Which of the enzymatic production or the intracellular stores mainly contributes to the physiological release of H₂S has not been determined.

H₂S-Sensitive Probes

The current methods to detect H₂S, such as colorimetric assays, gas chromatography, and polarographic analysis, require its extraction from cells and tissues. Because the catabolism and extinction of H₂S are fast, it has been difficult to measure the concentrations of this molecule accurately. Although a thiol detective imaging probe had been made, it was unable to differentiate sulfide from thiols. Recently, several fluorescent probes specifically detect H₂S have been developed (38, 39, 50, 52, 55).

SF1 and SF2

Based on the reaction that azides can be reduced by H₂S to amine, the azide-caged rhodamine analogues, SF1 and SF2, which generate highly fluorescent rhodamine products, were prepared (38). In a 30-min reaction with H₂S, SF1 and SF2 produce 7- and 9-fold responses, respectively. Although both probes slightly respond to •NO, they do not respond to reactive oxygen species, reactive nitrogen species, and thiols such as glutathione and cysteine. SF1 shows greater responses to H₂S in cells compared to SF2, probably due to the higher lipophilicity and cellular retention of SF1 relative to SF2. The improvement in shorter response time may help applying these probes to the studies of physiological and pathological processes.

DNS-Az

Dansyl is well-known for its strong fluorescence and long emission wavelength. DNS-Az is dansyl azide which emits fluorescence when H₂S reduces sulfonyl azide into sulfonamide (50). The reaction is similar to that of SF1 and SF2 (38).

Twenty five μM H_2S induces a 40-fold fluorescence enhancement. Although the detection is based on the reducing properties of H_2S , other possible reducing anions such as iodide, bromide, fluoride, bisulfate, and thiosulfate did not induce any responses. Although the selectivity of DNS-Az to H_2S compared to the other endogenous thiols, such as glutathione and cysteine, is not shown, its reaction with H_2S is fast, saturated in 5 min in 20 mM phosphate buffer in the presence of 0.05% Tween-20.

Probe 1

To selectively detect nucleophilic H_2S in the biological system, it is important to differentiate H_2S from other nucleophiles, especially thiols such as glutathione and cysteine. H_2S , which is a nonsubstituted thiol, can undergo nucleophilic reaction twice, while other thiols such as glutathione and cysteine are monosubstituted thiols that can only undergo nucleophilic reaction once. Probe 1 consists of a fluorophore, which is quenched by an attached structure containing an electrophilic component (39). H_2S reacts with the electrophilic component to form a free SH containing intermediate, which reacts with intrastructural ester group and causes cyclization to release the fluorophore.

Probe 1 reacts with 50 μM NaHS and the fluorescence intensity was increased by 55 to 70-fold, and the maximum intensity is reached in 1 h. Although probe 1 did not emit fluorescence in the presence of 50 μM glutathione and cysteine, its response to the endogenous levels of glutathione (mM order) was not shown. COS7 cells pre-incubated probe 1 emits strong fluorescence in the cells after the treatment of 250 μM NaHS.

SFP-1 and SFP-2

SFP-1 and SFP-2 consist of fluorophore and an aromatic framework with acrylate methyl ester and aldehyde ortho to each other (52). The aldehyde group reacts with sulfide to form a hemithioacetal intermediate with an exposed thiol, which reacts with the acrylate to yield a thioacetal. The reaction is similar to that of probe 1 (39). This reaction tunes photo-induced electron transfer of the aromatic structure, leading to the activation of fluorophore.

SFP-1 responds to 10–50 μM Na_2S and increases the intensity of fluorescence more than 10 times in 60 min in PBS buffer (pH 7.4). Although it also responds to the most abundant thiols in mammalian cells, glutathione and cysteine, the interference with H_2S can be excluded by selecting emission wave length. SFP-2, which has another fluorophore, showed more than 13-fold increase in the fluorescence intensity and more selective to Na_2S than to cysteine or glutathione. Although both probes slightly emit fluorescence, even without exogenously applied Na_2S in cultured HeLa cells, SFP-1 emits strong fluorescence in the presence of 100 μM Na_2S and SFP-1 with 200 μM Na_2S .

Hydrogen sulfide imaging probe-1

Azamacrocyclic rings form stable metal complexes with Cu^{2+} and the paramagnetic Cu^{2+} center has a pronounced quenching effect on fluorophores. Based on these characteristics of azamacrocyclic rings, hydrogen sulfide imaging probe-1 (HSip-1) was produced (55). H_2S binds to the Cu^{2+} center of HSip-1 to enhance fluorescence, while glutathione

does not cause any conformational change in the Cu^{2+} center or the increase in fluorescence intensity.

The fluorescence intensity is increased by 50-fold by 10 μM H_2S in HEPES buffer (pH 7.4) with the maximal intensity at 30 min, while no increase in fluorescence intensity is induced by 10 mM glutathione. HSip-1 has a high selectivity over other thiols such as 1 mM cysteine, homocysteine, and 2-mercaptoethanol, and 100 μM dithiothreitol (DTT), inorganic sulfur compounds, and a reducing compound. It does not respond to reactive oxygen species or reactive nitrogen species. HSip-1 DA, diacetylated HSip-1, is membrane permeable. HeLa cells emit fluorescence upon addition to 200 μM Na_2S to DMEM medium.

Production of H_2S

CBS

H_2S can be produced by three enzymes, CBS, CSE, and 3MST, along with cysteine aminotransferase (CAT), which is identical to aspartate aminotransferase (4, 7, 41). CBS produces H_2S from cysteine via a β -elimination reaction, and more efficiently via a β -replacement reaction in which cysteine is condensed with homocysteine (8). The plasma concentrations of homocysteine in Alzheimer's disease, heart attack, and stroke are elevated. The β -replacement reaction provides an explanation for the finding that pharmacological doses of N-acetylcysteine can lower plasma homocysteine concentrations, for N-acetylcysteine passes through plasma membrane to cytoplasm and is converted to cysteine, which is condensed by CBS with homocysteine to produce H_2S (8).

In the brain, astrocytes express CBS, which is intensively localized to cerebellar molecular layer and hippocampal dentate gyrus (16, 23) (Fig. 3). The specificity of our affinity-purified antibody against CBS was examined using Western blot analysis and immunohistochemistry by comparing CBS knockout and wild-type mice. The expression of CBS changes during development: it is expressed in neuroepithelial cells in

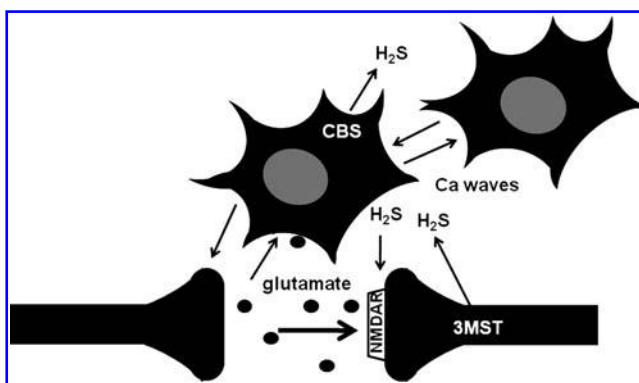


FIG. 3. CBS is localized to astrocytes and 3MST to neurons in the brain. There is a reciprocal interaction between neurons and astrocytes. Neuronal excitation induces Ca^{2+} influx in astrocytes, which in turn regulate synaptic activity. H_2S produced by CBS in astrocytes may be involved in the induction of Ca^{2+} waves, which propagate to surrounding astrocytes and contribute to the signaling between astrocytes. H_2S generated by CBS in astrocytes and by 3MST in neurons may be involved in enhancing the activity of NMDA receptors and the synaptic activity.

the ventricular zone at early developmental stages and in radial cells and astrocytes in the late embryonic and neonatal periods (16). The level of CBS is increased in reactive astrocytes, which mediate neuronal recovery in the injured brain, and is induced by epidermal growth factor (EGF), transforming growth factor (TGF)- α , and cyclic adenosine monophosphate (cAMP). Its levels are also increased in the hippocampus after kainic acid-induced seizures.

The cerebellum of CBS-knockout mice is remarkably smaller than that of wild-type animals (16). The abnormalities are not observed at birth but become apparent around postnatal day 8. Although the structural organization is normal, there was a striking reduction in the thickness of the molecular- and granular-layers with stunted Purkinje dendrites.

CBS is localized to human chromosome 21 (80). Because of trisomy 21 in Down syndrome (DS), CBS levels in patients are predicted to be 1.5-fold greater than those in normal individuals. However, the expression of CBS mRNA is 12 times greater in myeloblasts of DS children than in those of normal individuals, and the levels of CBS in DS patients are 2.4–2.9 times greater than those in age-matched normal individuals (23, 64). Individuals with DS almost invariably develop an Alzheimer type of dementia (AD). CBS is localized to astrocytes and the surrounding senile plaques in the brains of DS patients with AD. The overexpression of CBS may cause a developmental abnormality in cognition in DS children and that may lead to AD in DS adults (23). Overproduction of H₂S may cause the mental disturbance in DS. Another example of brain damage caused by elevated concentrations of H₂S is ethylmalonic encephalopathy, in which the gene that encodes sulfur dioxygenase (ETHE1) is mutated. Defects in sulfur dioxygenase, which normally metabolizes H₂S, lead to elevated concentrations of H₂S (66).

CSE

CSE can facilitate the production of H₂S from cysteine and homocysteine. Under normal conditions, α,β -elimination of cysteine generates H₂S. However, with high concentrations of homocysteine because of homocysteinemia, the α,γ -elimination and γ -replacement reactions of homocysteine become dominant for H₂S production by CSE (10). We could not detect CSE in the brain, and the activity of CSE is 100 times less than that in the liver (1, 17, 26). Although there are reports that show the existence of CSE activity in the brain, this may be due to the presence of a structurally unrelated protein that nonetheless exhibits CSE activity (72).

CSE is proposed to play a major role in vascular smooth muscle, and is mainly localized to smooth muscle in rodents (22, 82). However, CSE localization was recently reported in mouse, human, and bovine endothelium and cultures of human and bovine endothelium (81), although this finding is controversial (49, 57, 81, 82). The lysates of rat endothelium produce H₂S in the presence of cysteine and α -ketoglutarate, but there is no production of H₂S in the absence of α -ketoglutarate (57). Since CSE does not require α -ketoglutarate, this suggests that the other two H₂S-producing enzymes, 3MST and CAT, produce H₂S in rat endothelium. In bovine species, 3MST and CBS are localized to the endothelium, but CSE is not found (49).

Independent studies with CSE-knockout mice show that the involvement of CSE in the regulation of blood pressure is

also controversial (25, 81). The observation that blood pressure is unaffected by loss of CSE could be explained by at least two reasons. First, it is possible that 3MST and CAT may themselves be involved in the regulation of blood pressure. Alternatively, increased generation of H₂S by 3MST and CAT may compensate for loss of CSE in the knockout mice.

3MST with CAT

The production of H₂S from 3-mercaptopyruvate (3MP) by 3MST was initially measured at pH 7.4 (41). However, maximum production was found to occur under alkaline conditions (60, 62). CBS and CSE are localized to the cytoplasm, while 3MST is mainly localized to the mitochondrial matrix, with a pH of approximately 8 (45). 3MST is localized to neurons in the brain and retina (43, 58). Neither CBS nor CSE was found in the retina.

3MP is produced by CAT from cysteine and α -ketoglutarate (11, 70) (Fig. 4). The concentrations of cysteine in the cytosol are 0.15–0.25 mM and those in the mitochondria are 0.7–0.99 mM, which may be enough to produce the basal concentrations of H₂S (21, 63). In addition, glutathione, which is an intracellular store of cysteine, is present in 10–100 times greater concentrations in mammalian tissues than is cysteine, and can supply cysteine when the cells need it (63).

A reducing substance such as DTT is required for 3MST to produce H₂S. However, an endogenous substance corresponding to DTT has not been identified. We recently found that thioredoxin and DHLA associate with 3MST to release H₂S (42) (Fig. 4). Approximately 20 μ M thioredoxin exists in mitochondria, and it is 4 times more potent than DTT in enhancing H₂S production. The concentration of DHLA is approximately 40 μ M in the brain, and DHLA enhances the H₂S production as effectively as DTT. The redox potentials of dithiols such as DTT, DHLA, and 2 cysteine residues in the active site of thioredoxin are in the range of -0.26 to -0.33 V, while those of monothiols such as glutathione, cysteine, and

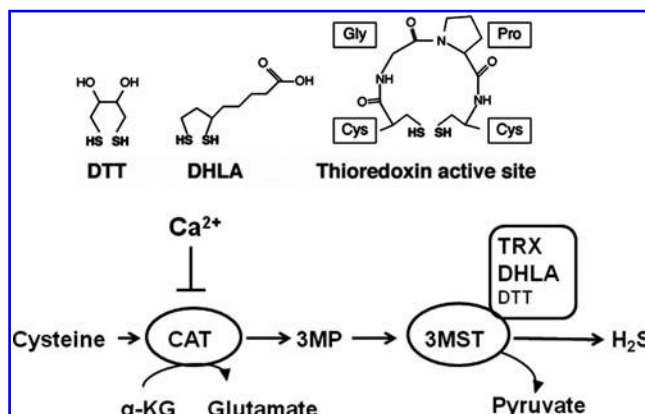


FIG. 4. Regulation of H₂S production by 3MST. Disulfides, such as thioredoxin (TRX) and dihydrolipoic acid (DHLA), are endogenous reducing substances, which can release H₂S from 3MST. The activity of CAT is regulated by Ca²⁺. In the absence of Ca²⁺, the production of H₂S by 3MST/CAT pathway from cysteine and α -ketoglutarate is the maximum but is suppressed by Ca²⁺ in a concentration-dependent manner.

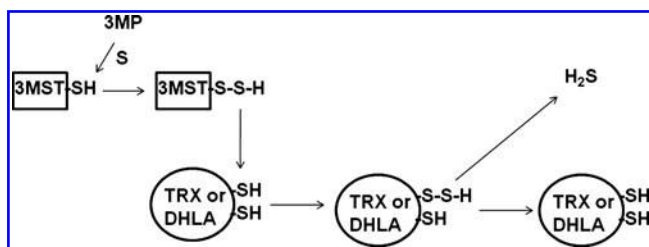


FIG. 5. A mechanism of releasing H_2S from 3MST in the presence of thioredoxin or DHLA. Thioredoxin and DHLA receive persulfide from 3MST. Thiol in thioredoxin and DHLA reduces persulfide to release H_2S .

CoA are from -0.22 to -0.35V (27). Those of nicotinamide adenine dinucleotide (NAD^+) and nicotinamide adenine dinucleotide phosphate (NADP^+) are around -0.32V . Because DTT, DHLA, and thioredoxin, whose two cysteine residues at the active site may function as dithiol, are able to release H_2S from 3MST, the dithiol structure but not the redox potential is a critical factor for H_2S release. A possible mechanism is that persulfide, which is produced at the active site cysteine of 3MST by receiving sulfur from 3-MP, is transferred to one of the thiols in thioredoxin or DHLA. The transferred persulfide, in turn, is reduced by another thiol in thioredoxin or DHLA to release H_2S (42, 75) (Fig. 5).

We recently found that production of H_2S from cysteine by the 3MST/CAT pathway is regulated by Ca^{2+} (43). The activity of CAT is suppressed by Ca^{2+} in a concentration-dependent manner. The production of H_2S is maximal in the absence of Ca^{2+} , and becomes completely suppressed at $2.9\text{ }\mu\text{M}$ Ca^{2+} . The range of intracellular Ca^{2+} shifts to the lower concentrations in retinal photoreceptor cells (10 nM – 600 nM) compared to other types of cells in which the intracellular concentrations are between 100 nM and $1\text{--}2\text{ }\mu\text{M}$. When retinal photoreceptor cells are exposed to light, the intracellular concentrations of Ca^{2+} are decreased to 10 nM . Intracellular Ca^{2+} concentrations are increased to 600 nM in darkness. In this range of Ca^{2+} concentration, production of H_2S by the 3MST/CAT pathway is dramatically changed. Calmodulin or its inhibitor, W-7, does not change the effect of Ca^{2+} on H_2S production, suggesting that calmodulin is not involved in the regulation of H_2S production by the 3MST/CAT pathway (43).

Both H_2S and DHLA induce nuclear factor erythroid 2-related factor 2 (Nrf 2), which increases the expression of thioredoxin and thioredoxin reductase (6, 59). Since thioredoxin and DHLA enhances the activity of 3MST to produce H_2S , both substances may also promote H_2S production through Nrf2 induction.

Modification of the Activity by Sulfhydrylation

Modification of the activity of enzymes and receptors is brought about by reversible covalent reactions, including phosphorylation and adenosine diphosphate (ADP)-ribosylation. The reversible incorporation of sulfur into proteins has been considered as a further example of regulation by covalent modification (Fig. 6). Kato et al. demonstrated that serine dehydratase was inhibited by CSE in the presence of cysteine (29). The suppression of the enzyme activity is reversible by glutathione or DTT, suggesting the formation of a

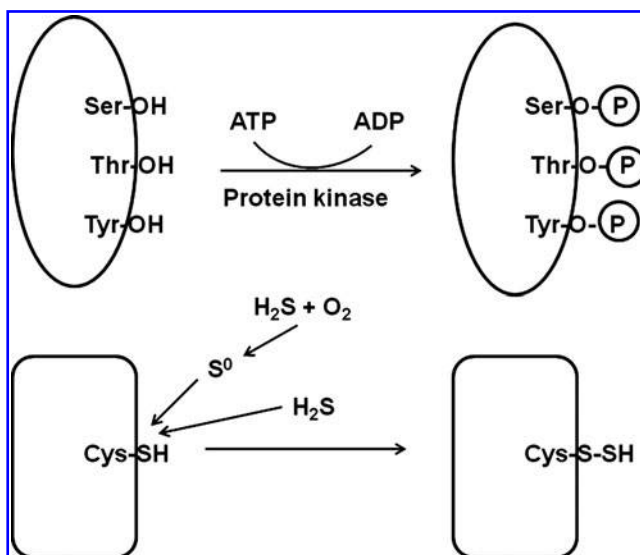


FIG. 6. Modulation of enzyme activity by phosphorylation and sulfurhydrylation. The reversible incorporation of sulfur into proteins may modulate protein function as phosphorylation does on proteins. H_2S may directly induce sulfurhydrylation. Alternatively, H_2S may react with oxygen to form S^0 that adds to sulfhydryl groups of proteins.

hydropersulfide group (sulfurhydrylation) in the enzyme. Enzymes inactivated by the same mechanism include 3-hydroxybutyrate dehydrogenase, alcohol dehydrogenase, and ornithine decarboxylase (67). Solomon Snyder and his colleagues recently demonstrated that H_2S induces sulfurhydrylation to enhance the activity of proteins, including glyceraldehyde 3-phosphate dehydrogenase (GAPDH), β -tubulin, and actin (19, 44). H_2S increases the activity of GAPDH by 7-fold, and DTT reverses GAPDH activity induced by H_2S . GAPDH is activated in human embryonic kidney (HEK) 293 cells expressing CSE, suggesting that endogenously produced H_2S induces sulfurhydrylation of GAPDH. However, further studies are required to determine the function of sulfurhydrylated GAPDH.

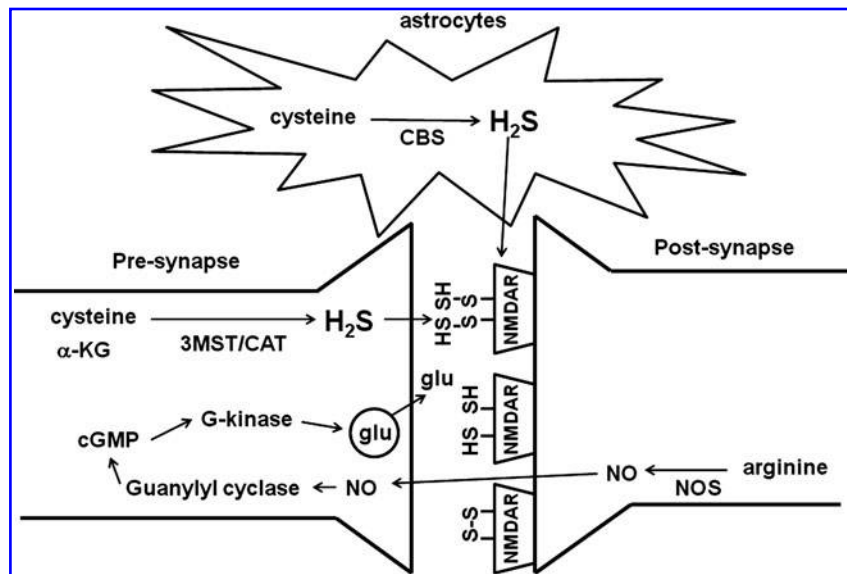
John Toohey proposed another mechanism for sulfurhydrylation. Some of H_2S reacts with oxygen to form S^0 that adds to sulfhydryl groups of proteins, forming a persulfide, and to disulfide groups, forming a trisulfide, though it is a generalized chemical mechanism and not site-specific (68).

Signaling Molecule

Synaptic modulation

The victims of H_2S poisoning suffer coordination and psychiatric disturbances (53). Based on these observations, the toxic effect of H_2S on the nervous system has been intensively studied. H_2S increases the levels of neurotransmitters γ -amino butyric acid (GABA), glutamate, serotonin, dopamine, epinephrine, and norepinephrine (33). The levels of serotonin and catecholamine are increased by inhibition of monoamine oxidase (MAO) (74). H_2S also enhances the effect of acetylcholine by suppressing the activity of its degradation enzyme, acetylcholinesterase. Excitatory postsynaptic potentials are suppressed by high concentrations of H_2S , indicating the suppression of synaptic transmission by H_2S (1). The effect

FIG. 7. The mechanisms for the induction of LTP. H₂S and nitric oxide (●NO) facilitate the induction of hippocampal long-term potentiation (LTP) with different mechanisms. ●NO, which is produced from arginine, diffuses to pre-synapse to activate guanylyl cyclase, leading to the production of cyclic GMP that activates G-kinase to increase the release of a neurotransmitter glutamate. Although H₂S does not have any effect on guanylyl cyclase, it enhances the activity of NMDA receptors.



may explain the temporary memory loss occurred following H₂S exposure.

H₂S facilitates the induction of hippocampal long-term potentiation (LTP) similarly to nitric oxide (●NO), albeit by a different mechanism (1, 48) (Fig. 7). ●NO, which is produced from arginine by ●NO synthetase at postsynapses, diffuses to presynapses to activate guanylyl cyclase. This leads to production of cyclic GMP, which in turn activates G-kinase to increase the release of a neurotransmitter glutamate. Carbon monoxide (CO) facilitates the induction of LTP in a similar fashion to ●NO, and both ●NO and CO are called retrograde neurotransmitters (48). In contrast H₂S does not have any effect on guanylyl cyclase (1). H₂S enhances the activity of NMDA receptors, which is known to be required for the induction of LTP. Disulfide bonds play a role in modulating the function of NMDA receptors (2). DTT reduces disulfide bonds to generate thiols in NMDA receptors and facilitates the induction of LTP. H₂S further enhances the induction of LTP even after the facilitation induced by DTT (1). These observations suggest that H₂S may be involved at two steps in the modulation of NMDA receptor activity. First, as observed for DTT, H₂S reduces disulfide bonds to generate thiols and enhance the activity of NMDA receptors. Second, H₂S potentiates the activity even further by binding to thiols of receptors and generating persulfide, as suggested by Snyder and Toohy (19, 24, 68).

Another possible mechanism for the modulation of synaptic activity observed in the facilitation of LTP induction may involve astrocytes. Astrocytes were thought to be quiescent cells whose primary role was to support neurons. However, they were recently found to have neurotransmitter-responsive receptors themselves. Neuronal excitation activates astrocytes, and these cells in turn modulate synaptic activity (12). In neurons, H₂S enhances the responses of NMDA receptors to glutamate, while H₂S induces Ca²⁺ influx in astrocytes, which enhances the synaptic activity (1, 47). The effects of H₂S on both neurons and astrocytes may therefore explain its role in the induction of LTP. Astrocyte responses are suppressed by La³⁺, Gd³⁺, and ruthenium red, which are known as transient receptor potential (TRP)

channel blockers, although the affected channel has not been identified.

Glial fibrillary acidic protein (GFAP) negative-premature astrocytes do not respond to H₂S (69). GFAP positive-mature astrocytes and those induced to mature by treatment with leukemia inhibitory factor (LIF) respond well to H₂S. In contrast, reactive astrocytes induced by EGF, TGF- α , cAMP, and interleukin-1 β (IL-1 β) do not respond to H₂S. Reactive astrocytes were induced by brain injuries, including trauma, neurodegenerative diseases, and viral infections. EGF is secreted by microglia and TGF- α is synthesized by injured neurons (28). EGF receptors are upregulated in reactive astrocytes after brain injury, and IL-1 β is produced by reactive astrocytes. Astrocytes may exchange signals with neurons under normal conditions, but once reactive astrocytes are induced by brain injury, they may stop responding to H₂S to prevent transmitting inappropriate signals (Fig. 8).

Regulation of the activity of retinal neurons

The center-surround organization of the receptive field is one of the most important characteristics of retinal neurons. The negative feedback from horizontal cells to photoreceptor cells plays a key role in the center-surround organization. The negative feedback has been proposed to occur when Ca²⁺ channels of photoreceptor cells are suppressed by H⁺ released by vacuolar-type H⁺-ATPase present on horizontal cells. When retinal photoreceptor cells are exposed to light, cyclic GMP-gated channels are closed and the intracellular concentrations of Ca²⁺ are decreased. This results in a decrease in the release of the neurotransmitter glutamate to horizontal cells. Without the activation of glutamate receptors, the intracellular concentration of Ca²⁺ is maintained at a low level. Low Ca²⁺ concentrations in both photoreceptor and horizontal cells enhances the activity of 3MST/CAT to produce H₂S, which activates vacuolar-type H⁺-ATPase on horizontal cells (43). H⁺ suppresses Ca²⁺ channels on photoreceptor cell membrane. H₂S suppresses Ca²⁺ influx and regulates the negative feedback from horizontal cells to photoreceptor cells.

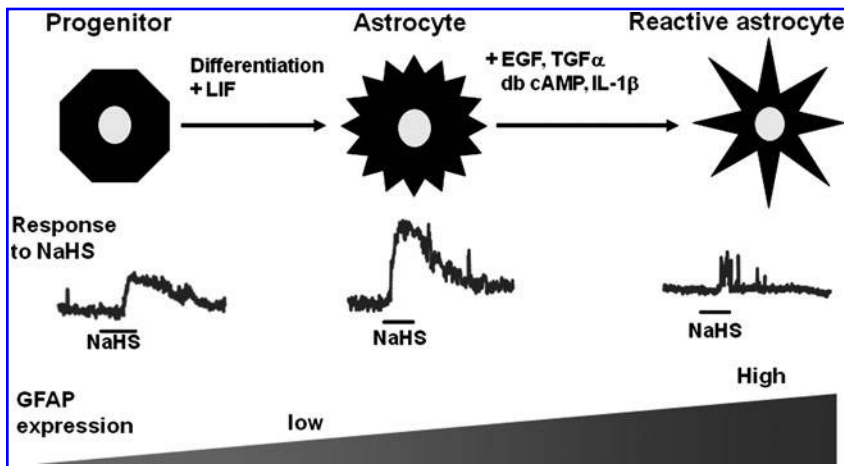


FIG. 8. The sensitivity of astrocytes to H_2S depends on the state of maturation and reactivity. GFAP positive-mature astrocytes and those maturation-induced by leukemia inhibitory factor (LIF) respond well to H_2S . In contrast, premature astrocytes as well as reactive astrocytes induced by EGF, $TGF\alpha$, cAMP, and interleukin- 1β do not respond to H_2S .

Cytoprotection

Because H_2S is a well-known toxic gas, the concept that it may also have a cytoprotective effect is paradoxical. We and others found that H_2S protects neurons from oxidative stress by reinstating glutathione levels decreased by oxidative stress and by scavenging reactive oxygen species (30–32, 76, 77). Primary cultures of immature neurons, which express cystine/cysteine transporters but do not express ionotropic glutamate receptors, are widely used as a model for oxidative stress (61). The model enabled us to study oxidative stress specifically without the involvement of excitotoxicity. Similarly, even 1 mM NaHS is not toxic to a neuroblastoma cell line, SH-SY5Y, which does not express NMDA receptors (76, 77). In contrast, primary cortical neurons, which were cultured for several days and express ionotropic glutamate receptors, were killed by NaHS, and cell death was inhibited by glutamate receptor antagonists, MK801, APV, and CNQX (9). Together, these results suggest the neuroprotective effect of H_2S is specific to oxidative stress, but not to excitotoxicity.

Under physiological conditions, H_2S dissociates to H^+ and HS^- and at equilibrium approximately 20% remains as undissociated H_2S gas, which can evaporate from the medium. Approximately one-third of total H_2S , which includes undissociated H_2S gas and dissociated HS^- , remains after 15 min of application, and more than 90% is lost from the medium of culture dishes by 30 min (30). After 1 h of application of NaHS, almost all of $H_2S + HS^-$ has evaporated out of culture dishes. Therefore, the effect of H_2S observed in culture dishes following a single application must be rapid and sustainable. It is possible that H_2S is bound to proteins as sulfane sulfur and in this manner can regulate their function (24, 68). Under oxidative stress caused by high concentrations of glutamate, neuronal cells started dying 8 h after exposure and only around 10% of cells survived after 16 h. Simultaneous application of glutamate and NaHS delayed the onset of cell death to 12 h and a rate of cell survival is much greater than that without NaHS. Because H_2S evaporates in a short time period, this means that the prolonged H_2S -dependent resistance to oxidative stress is induced by only a transient exposure.

H_2S can also protect cells against oxidative stress when given after the original insult. For example, the application of NaHS 4 h after induction of oxidative stress, still provides significant protection compared to control. This is true despite

the initial activation of a cell death cascade. This property of H_2S is important for its therapeutic application (51).

Increase in the levels of glutathione

H_2S increases the transport of cystine and cysteine by enhancing the activity of X_{CT}^- cystine/glutamate antiporters and $X_{A,G}^-$ excitatory amino acid transporters to increase the substrate concentrations for glutathione production. H_2S also enhances the activity of γ -glutamyl cysteine synthetase, a limiting enzyme for glutathione production. By these integrated effects H_2S increases the levels of glutathione in cells, and the effects are more profound when cells are under the conditions of oxidative stress (31, 32).

Various types of proapoptotic signals converge on mitochondria, and mitochondrial dysfunction causes numerous neurodegenerative diseases (37). 3MST and CAT are localized to mitochondria, which are the major organelles implicated in production of reactive oxygen species. H_2S produced in mitochondria may directly suppress oxidative stress in this organelle. For example, Neuro2a cells expressing 3MST and CAT show significant resistance to oxidative stress (31).

H_2S protects the embryonic brain from ischemia-reperfusion injury (31). Reperfusion after ischemia caused by stopping the blood flow from mother to fetus caused maceration of all the fetal brains, which contained only 24% glutathione of a control, but by the administration of sodium hydrosulfide (NaHS) to mother at the ischemia, 75% of fetal brains were not macerated and contained 90% of the normal glutathione level. The H_2S -producing enzymes that are also involved in the increase in the levels of glutathione have not been identified. The involvement of CSE in this process is controversial in two studies with CSE-knockout mice (25, 81).

Suppression of Ca^{2+} influx

The retina is susceptible to oxidative stress because of its high consumption of oxygen and daily exposure to light. Excessive light exposure leads to photoreceptor degeneration in the retina. Photoreceptor cell death is an irreversible injury that is caused by various factors such as reactive oxygen species (ROS) and elevated intracellular concentrations of Ca^{2+} . H_2S suppresses the elevation of Ca^{2+} in the photoreceptor cells by activating vacuolar-type H^+ ATPase, which

suppresses L-type Ca²⁺ channels, and maintains Ca²⁺ homeostasis (43). Intracellular concentrations of Ca²⁺ are increased during photoreceptor apoptosis, but the L-type Ca²⁺ channel blocker diltiazem prevents light-induced photoreceptor cell death. The lack of the vacuolar-type H⁺ ATPase α 3 subunit causes retinal degeneration. The failure of Ca²⁺ homeostasis due to excessive light may cause retinal cell degeneration. The regulation of Ca²⁺ and cytoprotective effect of endogenous H₂S may fail when photoreceptor cells are exposed to excessive light. Even under such conditions, the administration of H₂S protects cells from light-induced degeneration. Many cells in the outer nuclear layer, in which rod inner segments are located, became TUNEL-positive following exposure to light. The administration of 0.4375 μ mol NaHS/kg weight of mice decreased the number of TUNEL-positive cells by approximately 80% relative to a control. Light exposure also produces a lot of cells containing 8-hydroxy-2-deoxyguanosine (8-OHdG), which is a product of DNA damaged by ROS, in the outer nuclear layer. In contrast, the number of cells positive to 8-OHdG is decreased in NaHS-treated mice. H₂S protects photoreceptor cells from light-induced retinal degeneration and oxidative stress.

Trophic effect

A treatment of primary cultures of neurons with NaHS increases the reduction of tetrazolium salts, which are reduced by cellular activity to form intensely colored formazans and have often been used to quantitate the number of living cells (71). A similar effect is observed in neuroblastoma Neuro2a cells, but it is not observed in other cell types such as primary cultures of astrocytes, COS-7 and NIH3T3 fibroblast cell lines. H₂S enhances metabolic activity specifically in neurons and the neuronal cell line. Other reducing agents, glutathione, DTT, α -tocopherol, and 2-mercaptoethanol do not enhance metabolic activity of neurons. The effect is increased in 60 min, and the maximal effect is maintained at least for 24 h after the application of NaHS. The increase of tetrazolium salt reduction in the absence of cells, even without washing out NaHS, was much less than that in the presence of cells with washing out of NaHS. This excludes the possibility that the reduction of tetrazolium salts is directly caused by reducing activity of H₂S, and suggests that H₂S stimulates a reductive intracellular environment.

It is well-known that H₂S inhibits cytochrome c oxidase and suppresses respiratory chain metabolic activity (71). The concentrations of H₂S, which reduce tetrazolium salts, do not inhibit cytochrome c oxidase. In contrast, azide, another inhibitor of cytochrome c oxidase, greatly inhibited cytochrome c oxidase but did not enhance the reduction of tetrazolium salts. NaHS increases the metabolic activity without affecting NADH/NAD⁺ ratio. The effects of NaHS are suppressed by the tyrosine kinase inhibitors, genistein and tyrphostin A23, suggesting that the effect may be mediated by phosphorylation of tyrosine and that H₂S may have a trophic effect in the brain. Proliferation is induced by H₂S in other cell types, such as in lung cells, endothelial cells, cardiac myocytes, and umbilical vein endothelial cells, while apoptosis is induced in smooth muscle cells, lung primary fibroblast cells, and periodontal ligament cells (3). The induction of proliferation or apoptosis by H₂S in these cells is regulated by transcription of genes of kinases and cell cycle proteins.

H₂S-Based Therapeutics

A tremendous amount of effort has been devoted into the development of H₂S based therapeutics. Na₂S is being tested in bypass surgery patients in Phase II clinical trial, and for kidney injury in Phase I clinical trial (51). Since Na₂S and NaHS release H₂S immediately after they are applied *in vivo*, compounds slowly release H₂S in a physiological manner have been developed. For example, GYY4137 inhibits LPS-induced lung and liver injury by decreasing neutrophil accumulation as well as the levels of the anti-inflammatory chemokine IL-10 (78). GYY4137 also protects human articular chondrocytes and trabecular bone-derived mesenchymal progenitor cells from oxidative stress.

Nonsteroidal anti-inflammatory drugs (NSAIDs) are commonly used medications but have side effects, mainly gastric ulceration and bleeding. An H₂S-releasing derivative of naproxen (ATB-346) significantly accelerates the ulcer healing process, probably due to increasing the expression of cyclooxygenase (COX)-2 and promoting angiogenesis (5). ATB-346 does not cause gastric damage in rats even at exceedingly high doses. Another H₂S-releasing derivative of mesalamine (ATB-429) has greater anti-inflammatory activity in animal model of colitis compared to parental mesalamine. ACS14 and ACS21 are derivatives of aspirin and salicylic acid, respectively. Both compounds reduce hypertension and lower plasma levels of thromboxane B2 and insulin induced by glutathione depletion (54). S-diclofenac (ACS-15) efficiently suppresses LPS-induced inflammation with less gastric toxicity than parental diclofenac. This drug protects the heart from the development of ischemia-reperfusion injury and suppresses progression of vascular injury, leading to the prevention of arterial thrombosis.

Levodopa (L-DOPA) is the most widely used therapy for Parkinson's disease. The H₂S-releasing derivatives of L-DOPA (ACS5, ACS48, ACS50, and ACS81) were developed that showed significant anti-inflammatory, antioxidant, and neuroprotective properties (35). H₂S released from these compounds suppresses monoamine oxidase and maintains the effective concentrations of dopamine at the synaptic cleft. Released H₂S also suppresses the release of neurotoxins such as tumor necrosis factor- α , IL-6, and •NO from stimulated microglia. These compounds may be candidates for future treatment of Parkinson's disease.

Conclusion and Perspective

The basal concentrations of H₂S have been found to be much lower than those reported in earlier studies (18, 24, 79). However, the concentrations of H₂S achieved following cell stimulation are not well understood. To address this problem, it is necessary to clarify the regulation of the activity of H₂S-producing enzymes. For example, the activity of CBS is enhanced by S-adenosyl methionine and the 3MST/CAT pathway is regulated by Ca²⁺. The physiological stimulations, which regulate the levels of S-adenosyl methionine and Ca²⁺, may help determine the mechanism underlying the physiological regulation of CBS and 3MST/CAT.

In addition to the release of H₂S from the producing enzymes, bound sulfane sulfur, which can be an intracellular store of H₂S, releases H₂S under physiological conditions. The mechanism for the release of H₂S from bound sulfane sulfur may also help understand the endogenous local

concentrations of H₂S in the activated cells. The release of H₂S from activated astrocytes has not been successfully measured in the current method (24). Recently, fluorescent probes for imaging H₂S have been developed (38, 39, 50, 52, 55). The probes are able to selectively measure H₂S in aqueous environments at physiological pH. Although the sensitivity and intensity of the probes needs further improvement, their responses are selective for H₂S and can be applied to live cells for imaging. This represents a significant breakthrough for the detection of H₂S and is an improvement over the currently applied methods.

H₂S has the potential to be developed into a therapeutic agent (51). Na₂S is already in a Phase II clinical trial designed to determine whether it can protect cardiac muscle from ischemia-reperfusion injury following bypass surgery. NSAIDs attached to the H₂S-releasing group do not have side effects, such as bleeding or ulcer of gastrointestinal tract, that the parental NSAIDs cause (5). L-Dopa attached to an H₂S-releasing group has additional therapeutic effects that L-Dopa does not have (35). It suppresses the release of inflammatory cytokines produced by microglia, a key cause of neurodegeneration observed in Parkinson's disease. These are only a few examples of many H₂S-based therapeutic applications. Under physiological conditions, H₂S may be locally released to function in a restricted area. In contrast, the administration of H₂S systemically increases its concentrations. Monitoring for unexpected side effects will clearly be required. However, once these issues are overcome, the promise of H₂S-based therapeutic compounds will be realized.

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There is no conflict of interest for any of the authors.

References

1. Abe K and Kimura H. The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 16: 1066–1071, 1996.
2. Aizenman E, Lipton DA, and Loring RH. Selective modulation of NMDA responses by reduction and oxidation. *Neuron* 2: 1257–1263, 1989.
3. Basker R and Bian J. Hydrogen sulfide gas has cell growth regulatory role. *Eur J Pharmacol* 656: 5–9, 2011.
4. Braunstein AE, Goryachenkova EV, Tolosa EA, Willhardt IH, and Yefremova LL. Specificity and some other properties of liver serine sulphydrase: Evidence for its identity with cystathionine β -synthase. *Biochim Biophys Acta* 242: 247–260, 1971.
5. Caliendo G, Cirino G, Santagada V, and Wallace JL. Synthesis and biological effects of hydrogen sulfide (H₂S): Development of H₂S-releasing drugs as pharmaceuticals. *J Med Chem* 53: 6275–6286, 2010.
6. Calvert JW, Jha S, Gundewar S, Elrod JW, Ramachandran A, Pattillo CB, Kevil CG, and Lefer DJ. Hydrogen sulfide mediates cardioprotection through Nrf2 signaling. *Circ Res* 105: 365–374, 2009.
7. Cavallini D, Mondovi B, De Marco C, and Scioscia-Santoro A. The mechanism of desulphydration of cysteine. *Enzymologia* 24: 253–266, 1962.
8. Chen X, Jhee KH, and Kruger WD. Production of the neuromodulator H₂S by cystathionine β -synthase via the condensation of cysteine and homocysteine. *J Biol Chem* 279: 52082–52086, 2004.
9. Cheung NS, Peng ZF, Chen MJ, and Moore PK. Hydrogen sulfide induced neuronal death occurs via glutamate receptor and is associated with calpain activation and lysosomal rupture in mouse primary cortical neurons. *Neuropharmacology* 53: 505–514, 2007.
10. Chiku T, Padovani D, Zhu W, Singh S, Vitvitsky V, and Banerjee R. H₂S biogenesis by human cystathionine γ -lyase leads to the novel sulfur metabolites lanthionine and homolanthionine and is responsive to the grade of hyperhomocysteinemia. *J Biol Chem* 284: 11601–11612, 2009.
11. Cooper AJL. Biochemistry of sulfur-containing amino acids. *Annu Rev Biochem* 52: 187–222, 1983.
12. Dani JW, Chernjavsky A, and Smith SJ. Neuronal activity triggers calcium waves in hippocampal astrocyte networks. *Neuron* 8: 429–440, 1992.
13. Dello Russo C, Tringali G, Ragazzoni E, Maggiano N, Menini E, Vairano M, Preziosi P, and Navarra P. Evidence that hydrogen sulphide can modulate hypothalamo-pituitary-adrenal axis function: *In vitro* and *in vivo* studies in the rat. *J Neuroendocrinol* 12: 225–233, 2000.
14. Dombkowski RA, Russell MJ, and Olson KR. Hydrogen sulfide as an endogenous regulator of vascular smooth muscle tone in trout. *Am J Physiol Regul Integr Comp Physiol* 286: R678–685, 2004.
15. Elrod JW, Calvert JW, Morrison J, Doeller JE, Kraus DW, Tao L, Jiao X, Scalia R, Kiss L, Szabo C, et al. Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function. *Proc Natl Acad Sci USA* 104: 15560–15565, 2007.
16. Enokido Y, Suzuki E, Iwasawa K, Namekata K, Okazawa H, and Kimura H. Cystathionine β -synthase, a key enzyme for homocysteine metabolism, is preferentially expressed in the radial glia/astrocyte lineage of developing mouse CNS. *FASEB J* 19: 1854–1856, 2005.
17. Finkelstein JD. Methionine metabolism in mammals. *J Nutr Biochem* 1: 228–237, 1990.
18. Furne J, Saeed A, and Levitt MD. Whole tissue hydrogen sulfide concentrations are orders of magnitude lower than presently accepted values. *Am J Physiol Regul Integr Comp Physiol* 295: R1479–R1498, 2008.
19. Gadalla MM and Snyder SH. Hydrogen sulfide as a gasotransmitter. *J Neurochem* 113: 14–26, 2010.
20. Goodwin LR, Francom D, Dieken FP, Taylor JD, Warenycia MW, Reiffenstein RJ, and Dowling G. Determination of sulfide in brain tissue by gas dialysis/ion chromatography: Postmortem studies and two case reports. *J Anal Toxicol* 13: 105–109, 1989.
21. Griffith OW. Biologic and pharmacologic regulation of mammalian glutathione synthesis. *Free Radic Biol Med* 27: 922–935, 1999.
22. Hosoki R, Matsuki N, and Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant

- in synergy with nitric oxide. *Biochem Biophys Res Commun* 237: 527–531, 1997.
23. Ichinohe A, Kanaumi T, Takashima S, Enokido Y, Naai Y, and Kimura H. Cystathionine beta-synthase is enriched in the brains of Down's patients. *Biochem Biophys Res Commun* 338: 1547–1550, 2005.
24. Ishigami M, Hiraki K, Umemura K, Ogasawara Y, Ishii K, and Kimura H. A source of hydrogen sulfide and a mechanism of its release in the brain. *Antioxid Redox Signal* 11: 205–214, 2009.
25. Ishii I, Akahoshi N, Yamada H, Nakano S, Izumi T, and Suematsu M. Cystathionine γ -lyase-deficient mice require dietary cysteine to protect against acute lethal myopathy and oxidative injury. *J Biol Chem* 285: 26358–26368, 2010.
26. Ishii I, Akahoshi N, Yu X-N, Kobayashi Y, Namekata K, Komaki G, and Kimura H. Murine cystathionine γ -lyase: Complete cDNA and genomic sequences, promoter activity, tissue distribution and developmental expression. *Biochem J* 381: 113–123, 2004.
27. Jocelyn PC. The standard redox potential of cysteine-cystine form the thiol-disulphide exchange reaction with glutathione and lipoic acid. *Eur J Biochem* 2: 327–331, 1967.
28. Junier MP, Coudrier M, Le Forestier N, Cadusseau J, Suzuki F, Peschanski M, and Dreyfus PA. Transforming growth factor alpha expression in degenerating motoneurons of the murine mutant wobbler: A neuronal signal for astrogliosis? *J Neurosci* 14: 4206–4216, 1994.
29. Kato A, Ogura M, and Suda M. Control mechanism in the rat liver enzyme system converting L-methionine to L-cystine. 3. Noncompetitive inhibition of cystathionine synthetase-serine dehydratase by elemental sulfur and competitive inhibition of cystathionine-homoserine dehydratase by L-cysteine and L-cystine. *J Biochem* 59: 40–48, 1966.
30. Kimura Y, Dargusch R, Schubert D, and Kimura H. Hydrogen sulfide protects HT22 neuronal cells from oxidative stress. *Antioxid Redox Signal* 8: 661–670, 2006.
31. Kimura Y, Goto Y-I, and Kimura H. Hydrogen sulfide increases glutathione production and suppresses oxidative stress in mitochondria. *Antioxid Redox Signal* 12: 1–13, 2010.
32. Kimura Y and Kimura H. Hydrogen sulfide protects neurons from oxidative stress. *FASEB J* 18: 1165–1167, 2004.
33. Kombian SB, Warenycia MW, Mele FG, and Reiffenstein RJ. Effects of acute intoxication with hydrogen sulfide on central amino acid transmitter systems. *NeuroToxicology* 9: 587–596, 1988.
34. Lee JK, Kozono D, Remis J, Kitagawa Y, Agre P, and Stroud RM. Structural basis for conductance by the archaeal aquaporin AsqpM at 1.68 Å. *Proc Natl Acad Sci USA* 102: 18932–18937, 2005.
35. Lee M, Tazzari V, Glustarini D, Rossi R, Sparatore A, Soldato PD, McGeer E, and McGeer PL. Effects of hydrogen sulfide-releasing L-DOPA derivatives on glial activation. *J Biol Chem* 285: 17318–17328, 2010.
36. Levitt MD, Abdel-Rehim MS, and Furne J. Free and acid-labile hydrogen sulfide concentrations in mouse tissues: Anomalous high free hydrogen sulfide in aortic tissue. *Antioxid Redox Signal* 15: 373–378, 2011.
37. Lin MT and Beal MF. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 443: 787–795, 2006.
38. Lippert AR, New EJ, and Chang CJ. Re-based fluorescent probes for selective imaging of hydrogen sulfide in living cells. *J Am Chem Soc* 133: 10078–10080, 2010.
39. Liu C, Pan J, Li S, Zhao Y, Wu LY, Berkman CE, Whorton AR, and Xian M. Capture and visualization of hydrogen sulfide by a fluorescent probe. *Angew Chem Int Ed* 50: 10327–10329, 2011.
40. Mathai JC, Missner A, Kugler P, Saparov SM, Zeidel ML, Lee JK, and Pohl P. No facilitator required for membrane transport of hydrogen sulfide. *Proc Natl Acad Sci USA* 106: 16633–16638, 2009.
41. Meister A, Fraser PE, and Tice SV. Enzymatic desulfuration of β -mercaptopyruvate to pyruvate. *J Biol Chem* 206: 561–575, 1954.
42. Mikami Y, Shibuya N, Kimura Y, Nagahara N, Ogasawara Y, and Kimura H. Thioredoxin and dihydrolipoic acid are required for 3-mercaptopyruvate sulfurtransferase to produce hydrogen sulfide. *Biochem J* 439: 479–485, 2011.
43. Mikami Y, Shibuya N, Kimura Y, Nagahara N, Yamada M, and Kimura H. Hydrogen sulfide protects the retina from light-induced degeneration by the modulation of Ca²⁺ influx. *J Biol Chem* 286: 39379–39386, 2011.
44. Mustafa AK, Gadalla MM, Sen N, Kim S, Mu W, Gazi SK, Barrow RK, Yang G, Wang R, and Snyder SH. H₂S signals through protein S-sulfhydration. *Sci Signal* 2: ra72, 2009.
45. Nagahara N, Ito T, Kitamura H, and Nishino T. Tissue and subcellular distribution of mercaptopyruvate sulfurtransferase in the rat: Confocal laser fluorescence and immunoelectron microscopic studies combined with biochemical analysis. *Histochem J* 110: 243–250, 1998.
46. Nagahara N, Okazaki T, and Nishino T. Cytosolic mercaptopyruvate sulfurtransferase is evolutionarily related to mitochondrial rhodanese. Striking similarity in active site amino acid sequence and the increase in the mercaptopyruvate sulfurtransferase activity of rhodanese by site-directed mutagenesis. *J Biol Chem* 270: 16230–16235, 1995.
47. Nagai Y, Tsugane M, Oka J, and Kimura H. Hydrogen sulfide induces calcium waves in astrocytes. *FASEB J* 18: 557–559, 2004.
48. O'Dell TJ, Hawkins RD, Kandel ER, and Arancio O. Tests of the roles of two diffusible substances in long-term potentiation: Evidence for nitric oxide as a possible early retrograde messenger. *Proc Natl Acad Sci USA* 88: 11285–11289, 1991.
49. Olson KR, Whitfield NL, Bearden SE, Leger JS, Nilson E, Gao Y, and Maddeen JA. Hypoxic pulmonary vasodilation: A paradigm shift with a hydrogen sulfide mechanism. *Am J Physiol Regul Integr Comp Physiol* 298: R51–R60, 2010.
50. Peng H, Cheng Y, Dai C, King AL, Predmore BL, Lefer DJ, and Wang B. A fluorescent probe for fast and quantitative detection of hydrogen sulfide in blood. *Angew Chem Int Ed* 50: 9672–9675, 2011.
51. Predmore BL and Lefer DJ. Development of hydrogen sulfide-based therapeutics for cardiovascular disease. *J Cardiovasc Transl Res* 3: 487–498, 2010.
52. Qian Y, Karpus J, Klabil O, Zhang S-Y, Zhu H-L, Banerjee R, Zhao J and He C. Selective fluorescent probes for live-cell monitoring of sulphide. *Nat Commun* 2–495, 2011.
53. Reiffenstein RJ, Hulbert WC, and Roth SH. Toxicology of hydrogen sulfide. *Annu Rev Pharmacol Toxicol* 32: 109–134, 1992.
54. Rossoni G, Manfredi B, Tazzari V, Sparatore A, Trivulzio S, Del Soldato P, and Berti F. Activity of a new hydrogen sulfide-releasing aspirin (ACS14) on pathological cardiovascular alterations induced by glutathione depletion in rats. *Eur J Pharmacol* 648: 139–145, 2010.
55. Sasakura K, Hanaoka K, Shibuya N, Mikami Y, Kimura Y, Komatsu T, Ueno T, Terai T, Kimura H, and Nagano T.

- Development of a highly selective fluorescence probe for hydrogen sulfide. *J Am Chem Soc* 133: 18003–18005, 2011.
56. Savage JC and Gould DH. Determination of sulfide in brain tissue and rumen fluid by ion-interaction reversed-phase high-performance liquid chromatography. *J Chromatogr* 526: 540–545, 1990.
 57. Shibuya N, Mikami Y, Kimura Y, Nagahara N, and Kimura H. Vascular endothelium expresses 3-mercaptopyruvate sulfurtransferase and produces hydrogen sulfide. *J Biochem* 146: 623–626, 2009.
 58. Shibuya N, Tanaka M, Yoshida M, Ogasawara Y, Togawa T, Ishii K, and Kimura H. 3-Mercaptopyruvate sulfurtransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxid Redox Signal* 11: 703–714, 2009.
 59. Suh JH, Shenvi SV, Dixon BM, Liu H, Jaiswal AKI, Liu RM, and Hagen TM. Decline in transcriptional activity of Nrf2 causes age-related loss of glutathione synthesis, which is reversible with lipoic acid. *Proc Natl Acad Sci USA* 101: 3381–3386, 2004.
 60. Stipanuk MH and Beck PW. Characterization of the enzymic capacity for cysteine desulphhydration in liver and kidney of the rat. *Biochem J* 206: 267–277, 1982.
 61. Tan S, Schubert D, and Maher P. Oxytosis: A novel form of programmed cell death. *Cur Top Med Chem* 1: 497–506, 2001.
 62. Taniguchi T and Kimura T. Role of 3-mercaptopyruvate sulfurtransferase in the formation of the iron-sulfur chromophore of adrenal ferredoxin. *Biochim Biophys Acta* 364: 284–295, 1974.
 63. Tateishi N, Higashi T, Naruse A, Nakashima K, and Shiozaki H. Rat liver glutathione: Possible role as a reservoir of cysteine. *J Nutr* 107: 51–60, 1977.
 64. Taub JW, Huang X, Matherly LH, Stout ML, Buck SA, Massey GV, Becton DL, Chang MN, Weinstein HJ, and Ravindranath Y. Expression of chromosome 21-localized genes in acute myeloid leukemia: Differences between Down syndrome and non-Down syndrome blast cells and relationship to *in vitro* sensitivity to cytosine arabinoside and daunorubicin. *Blood* 94: 1393–1400, 1999.
 65. Teague B, Asiedu S, and Moore PK. The smooth muscle relaxant effect of hydrogen sulphide *in vitro*: Evidence for a physiological role to control intestinal contractility. *Br J Pharmacol* 137: 139–145, 2002.
 66. Tiranti V, Viscomi C, Hildebrandt T, Di Meo I, Mineri R, Tiveron C, Levitt MD, Prella A, Fagioli G, Rimoldi M, and Zeviani M. Loss of ETHE1, a mitochondrial dioxygenase, causes fatal sulfide toxicity in ethylmalonic encephalopathy. *Nat Med* 15: 200–205, 2009.
 67. Toohey JI. Sulphane sulphur in biological systems: A possible regulatory role. *Biochem J* 264: 625–632, 1989.
 68. Toohey JI. Sulfur signaling: Is the agent sulfide or sulfane? *Anal Biochem* 413: 1–7, 2011.
 69. Tsugane M, Nagai Y, Kimura Y, Oka J-I, and Kimura H. Differentiated astrocytes acquire sensitivity to hydrogen sulfide that is diminished by the transformation into reactive astrocytes. *Antioxid Redox Signal* 9: 257–269, 2007.
 70. Ubuka T, Umemura S, Yuasa S, Kinuta M, and Watanabe K. Purification and characterization of mitochondrial cysteine aminotransferase from rat liver. *Physiol Chem Physics* 10: 483–500, 1978.
 71. Umemura K and Kimura H. Hydrogen sulfide enhances reducing activity in neurons: Neurotropic role of H₂S in the brain? *Antioxid Redox Signal* 9: 2035–2041, 2007.
 72. Vitvitsky V, Tomas M, Ghorpade A, Gendelman HE, and Banerjee R. A functional transsulfuration pathway in the brain links to glutathione homeostasis. *J Biol Chem* 281: 35785–35793, 2006.
 73. Warenycia MW, Goodwin LR, Benishin CG, Reiffenstein RJ, Grancom DM, Taylor JD, and Dieken FP. Acute hydrogen sulfide poisoning. Demonstration of selective uptake of sulfide by the brainstem by measurement of brain sulfide levels. *Biochem Pharmacol* 38: 973–981, 1989.
 74. Warenycia MW, Smith KA, Blashko CS, Kombian SB, and Reiffenstein RJ. Monoamine oxidase inhibition as a sequel of hydrogen sulfide intoxication: Increases in brain catecholamine and 5-hydroxytryptamine. *Arch Toxicol* 63: 131–136, 1989.
 75. Westrop GD, Georg I, and Coombs GH. The mercaptopyruvate sulfurtransferase of *Trichomonas vaginalis* links cysteine catabolism to the production of thioredoxin persulfide. *J Biol Chem* 284: 33485–33494, 2009.
 76. Whiteman M, Armstrong JS, Chu SH, Jia-Ling S, Wong, BS, Hheung NS, Halliwell B, and Moore PK. The novel neuro-modulator hydrogen sulfide: An endogenous peroxynitrite 'scavenger'? *J Neurochem* 90: 765–768, 2004.
 77. Whiteman M, Cheung NS, Zhu YZ, Chu SH, Siau JL, Wong BS, Armstrong JS, and Moore PK. Hydrogen sulphide: A novel inhibitor of hypochlorous acid-mediated oxidative damage in the brain? *Biochem Biophys Res Commun* 326: 794–798, 2005.
 78. Whiteman M and Winyard PG. Hydrogen sulfide and inflammation: The good, the bad, the ugly and the promising. *Expert Rev Clin Pharmacol* 4: 13–32, 2011.
 79. Wintner EA, Deckwerth TL, Langston W, Bengtsson A, Leviten D, Hill P, Insko MA, Dumpit R, VandenEkar E, Toombs CF, and Szabo C. A monobromobimane-based assay to measure the pharmacokinetic profile of reactive sulphide species in blood. *Br J Pharmacol* 160: 941–957, 2010.
 80. Wisniewski KE, Wisniewski HM, and Wen GY. Occurrence of neuropathological changes and dementia of Alzheimer's disease in Down's syndrome. *Ann Neurol* 17: 278–282, 1985.
 81. Yang G, Wu L, Jiang B, Yang W, Qi J, Cao K, Meng Q, Mustafa AK, Mu W, Zhang S, et al. H₂S as a physiologic vasorelaxant: Hypertension in mice with deletion of cystathionine γ -lyase. *Science* 322: 587–590, 2008.
 82. Zhao W, Zhang J, Lu Y, and Wang R. The vasorelaxant effect of H₂S as a novel endogenous gaseous K_{ATP} channel opener. *EMBO J* 20: 6008–6016, 2001.

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Abbreviations Used

AD = Alzheimer type of dementia
ADP = adenosine diphosphate
ATP = adenosine triphosphate
cAMP = cyclic adenosine monophosphate
CAT = cysteine aminotransferase
CBS = cystathionine β -synthase
cGMP = cyclic guanosine monophosphate
CO = carbon monoxide
CRH = corticotropin-releasing hormone
CSE = cystathionine γ -lyase
DHLA = dihydrolipoic acid
DS = Down's syndrome
DTT = dithiothreitol
EDHF = endothelium-dependent hyperpolarizing factor
EDRF = endothelium-dependent relaxing factor
EGF = epidermal growth factor
GABA = γ -amino butyric acid
GAPDH = glyceraldehydes 3-phosphate dehydrogenase

GFAP = glial fibrillary acidic protein
HEK = human embryonic kidney
H₂S = hydrogen sulfide
IL = interleukin
K_{ATP} = ATP-dependent K⁺
L-Dopa = levodopa
LIF = leukemia inhibitory factor
LTP = long-term potentiation
MAO = monoamine oxidase
3MP = 3-mercaptopyruvate
3MST = 3-mercaptopyruvate sulfurtransferase
NADH = nicotinamide adenine dinucleotide
NADPH = nicotinamide adenine dinucleotide phosphate
NaHS = sodium hydrosulfide
NMDA = N-methyl D,L-aspartate
●NO = nitric oxide
Nrf2 = nuclear factor erythroid 2-related factor 2
NSAID = nonsteroidal anti-inflammatory drug
RNA = ribonucleic acid
TGF = transforming growth factor
TRP = transient receptor potential
TRX = thioredoxin

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